

EUPHORBIACEAE

FLAVONOIDS AND FATTY CONSTITUENTS OF *ADENOPELTIS*
COLLIGUAYA

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Plant. *Adenopeltis colliguaya* Bert. *Source.* Leaves collected in Rocoto, Concepcion, Chile in January (summer) and September (spring).

Extraction. (a) *Light petroleum.* Dried, powdered leaves (1.7 kg) extd. with light petroleum (60–80°) in a Soxhlet. After concn a dark green extract was obtained (74 g); a portion (30 g) in light petroleum was chromatographed on silica gel (350 g) and eluted with light petroleum (A) with light petroleum–benzene (B, C) and with benzene–EtOH (D).

(A) Recrystallized from benzene–MeOH afforded a waxy solid (99 mg). Preparative GLC afforded, as the major component, *nonacosane*,¹ m.p. 42–44°, ν_{\max} (Nujol) 1461, 1377, and 720 cm^{-1} . Its MS showed a parent ion at m/e 408 and a consistent fragmentation pattern. (B) Recrystallization from benzene afforded a solid (260 mg), m.p. 57–59°, ν_{\max} (KBr) 2938, 2858, 1735, 1465, and 1370 cm^{-1} , with a positive test to Brady's reagent and MS peaks at m/e 480, 465, 452, 424, 410, 409, 408, 396, 381, 218, 189, 157, 203, 101, 88, and 71. This ketonic fraction was not investigated further. (C). Recrystallization from light petroleum–MeOH gave a solid, m.p. 58–60°, ν_{\max} (KBr) 3390, 2933, 2865, 1471, 1379, 1058, 730, and 720 cm^{-1} . Its MS fragmentation pattern was consistent for a mixture of *tetratriacontanol*, *dotriacontanol*, and *triacontanol*, with the latter predominating.² (D). *Sitosterol* (190 mg), m.p. (MeOH) 138°, λ_{\max} (cyclohexane) 205 nm (end absorption), ν_{\max} (KBr) 3475, 1645 cm^{-1} . Its NMR spectrum and X-ray diffraction pattern were identical to those of an authentic sample. Its acetate also corresponded in props to an authentic sample.

(b) *Aqueous methanol extract.* The dried, powdered leaves (200 g) extd. with hot aq. MeOH (1:4), giving a gummy solid (125 g) on concn. Chromatography through silica gel, using benzene–EtOAc–EtOH mixtures, was followed by TLC on cellulose, using BAW as mobile phase,³ gave the following flavonoids (elution solvent in parentheses): *astragalin*⁴ (9:1, EtOAc–EtOH), acid hydrolysis⁵ gave kaempferol and glucose; *avicularin*⁶ (9:1,

¹ P. N. RYLANDER and S. MEYERSON, *J. Am. Chem. Soc.* **78**, 5799 (1956).

² R. A. FRIEDEL, J. L. SCHULTZ and A. G. SHARKEY, *Analyt. Chem.* **28**, 926 (1956).

³ J. B. HARBORNE, *J. Chromatog.* **2**, 581 (1959).

⁴ T. NAKABAYASHI, *J. Agric. Chem. Soc. Japan* **26**, 539 (1952).

⁵ J. B. HARBORNE, *Phytochem.* **4**, 107 (1965).

⁶ H. EL KHADAM and Y. S. MOHAMMED, *J. Chem. Soc.* 3320 (1952).

EtOAc-EtOH) (43 mg), $[\alpha]_D^{20} -161^\circ$ (c 1.0, EtOH), acid hydrolysis gave quercetin and arabinose; *quercitrin*⁷ (25 mg) (7:3, EtOAc-EtOH), m.p. 183–185°, acid hydrolysis gave quercetin and rhamnose; *meratin*⁸ (28 mg) (1:4, EtOAc-EtOH), m.p. 182–183°, acid hydrolysis afforded quercetin and glucose; *rutin*⁹ (33 mg) (EtOH), m.p. 178°, acid hydrolysis afforded quercetin, rhamnose and glucose. All the flavonoids were identified by direct comparison with authentic samples and by UV spectral analysis.

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⁷ T. A. GEISSMAN, *The Chemistry of Flavonoid Compounds*, p. 336, Macmillan, New York (1962); T. J. MABRY, K. R. MARKHAM and M. B. THOMAS, *The Systematic Identification of Flavonoids*, Springer-Verlag, New York (1970).

⁸ O. SCHINDLER, *Helv. Chim. Acta* **28**, 1157 (1945).

⁹ T. A. GEISSMAN, *The Chemistry of Flavonoid Compounds*, p. 338, Macmillan, New York (1962).

Key Word Index—*Adenopeltis colliguaya*; Euphorbiaceae; kaempferol and quercetin glycosides; sitosterol; hydrocarbons.

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TRITERPENOIDS AND STEROIDS OF *EUPHORBIA PILULIFERA*

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Plant. Euphorbia pilulifera. Previous work. Alkaloids from the latex.¹

Present work. Leaves and stems. The neutral fraction of hot hexane extract was chromatographed on an Al₂O₃ column. The following pentacyclic triterpenes and sterols were identified: taraxerol only in free form (m.p., acetate and ketone,² TLC, GLC and IR); taraxerone (m.p.,² TLC, GLC and IR). α - and β -Amyrin (10:1) were identified by GLC in the esterified form only. Campesterol (20%), stigmasterol (10%) and sitosterol (70%) occurred in both free and esterified forms (TLC and GLC) in mixtures. No euphol, euphorbol, tirucallol or other tetracyclic triterpenes could be detected in this plant.³

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¹ F. B. POWER and H. BROWNING, *Chem. Zentr.* **1**, 1824 (1913).

² J. SIMONSEN and W. C. J. ROSS, *The Terpenes*, Vol. IV, Cambridge Publishers (1957).

³ G. PONSINET and G. OURISSON, *Phytochem.* **7**, 89 (1969).

Key Word Index—*Euphorbia pilulifera*; Euphorbiaceae; taraxerone; taraxerol; α - and β -amyrin; steroids.